

Climatic controls of decomposition drive the global biogeography of forest–tree symbioses

B. S. Steidinger^{1,15}, T. W. Crowther^{2,15*}, J. Liang^{3,4,15*}, M. E. Van Nuland¹, G. D. A. Werner⁵, P. B. Reich^{6,7}, G. Nabuurs⁸, S. de-Miguel^{9,10}, M. Zhou³, N. Picard¹¹, B. Herault^{12,13}, X. Zhao⁴, C. Zhang⁴, D. Routh², GFBI consortium¹⁴ & K. G. Peay^{1*}

The identity of the dominant root-associated microbial symbionts in a forest determines the ability of trees to access limiting nutrients from atmospheric or soil pools^{1,2}, sequester carbon^{3,4} and withstand the effects of climate change^{5,6}. Characterizing the global distribution of these symbioses and identifying the factors that control this distribution are thus integral to understanding the present and future functioning of forest ecosystems. Here we generate a spatially explicit global map of the symbiotic status of forests, using a database of over 1.1 million forest inventory plots that collectively contain over 28,000 tree species. Our analyses indicate that climate variables—in particular, climatically controlled variation in the rate of decomposition—are the primary drivers of the global distribution of major symbioses. We estimate that ectomycorrhizal trees, which represent only 2% of all plant species⁷, constitute approximately 60% of tree stems on Earth. Ectomycorrhizal symbiosis dominates forests in which seasonally cold and dry climates inhibit decomposition, and is the predominant form of symbiosis at high latitudes and elevation. By contrast, arbuscular mycorrhizal trees dominate in aseasonal, warm tropical forests, and occur with ectomycorrhizal trees in temperate biomes in which seasonally warm-and-wet climates enhance decomposition. Continental transitions between forests dominated by ectomycorrhizal or arbuscular mycorrhizal trees occur relatively abruptly along climate-driven decomposition gradients; these transitions are probably caused by positive feedback effects between plants and microorganisms. Symbiotic nitrogen fixers—which are insensitive to climatic controls on decomposition (compared with mycorrhizal fungi)—are most abundant in arid biomes with alkaline soils and high maximum temperatures. The climatically driven global symbiosis gradient that we document provides a spatially explicit quantitative understanding of microbial symbioses at the global scale, and demonstrates the critical role of microbial mutualisms in shaping the distribution of plant species.

Microbial symbionts strongly influence the functioning of forest ecosystems. Root-associated microorganisms exploit inorganic, organic² and/or atmospheric forms of nutrients that enable plant growth¹, determine how trees respond to increased concentrations⁶ of CO₂, regulate the respiratory activity of soil microorganisms^{3,8} and affect plant species diversity by altering the strength of conspecific negative density dependence⁹. Despite the growing recognition of the importance of root symbioses for forest functioning^{1,6,10} and the potential to integrate symbiotic status into Earth system models that predict functional changes to the terrestrial biosphere¹⁰, we lack spatially explicit quantitative maps of root symbioses at the global scale. Quantitative maps of tree symbiotic states would link the biogeography of functional traits

of belowground microbial symbionts with their 3.1 trillion host trees¹¹, which are spread across Earth's forests, woodlands and savannahs.

The dominant guilds of tree root symbionts—arbuscular mycorrhizal fungi, ectomycorrhizal fungi, ericoid mycorrhizal fungi and nitrogen-fixing bacteria (N-fixers)—are all based on the exchange of plant photosynthate for limiting macronutrients. Arbuscular mycorrhizal symbiosis evolved nearly 500 million years ago, and ectomycorrhizal, ericoid mycorrhizal and N-fixer plant taxa have evolved multiple times from an arbuscular-mycorrhizal basal state. Plants that are involved in arbuscular mycorrhizal symbiosis comprise nearly 80% of all terrestrial plant species; these plants principally rely on arbuscular mycorrhizal fungi for enhancing mineral phosphorus uptake¹². In contrast to arbuscular mycorrhizal fungi, ectomycorrhizal fungi evolved from multiple lineages of saprotrophic ancestors and, as a result, some ectomycorrhizal fungi are capable of directly mobilizing organic sources of soil nutrients (particularly nitrogen)². Associations with ectomycorrhizal fungi—but not arbuscular mycorrhizal fungi—have previously been shown to enable trees to accelerate photosynthesis in response to increased concentrations of atmospheric CO₂ when soil nitrogen is limiting⁶, and to inhibit soil respiration by decomposer microorganisms^{3,8}. Because increased plant photosynthesis and decreased soil respiration both reduce atmospheric CO₂ concentrations, the ectomycorrhizal symbiosis is associated with buffering the Earth's climate against anthropogenic change.

In contrast to mycorrhizal fungi, which extract nutrients from the soil, symbiotic N-fixers (Rhizobia and Actinobacteria) convert atmospheric N₂ to plant-usable forms. Symbiotic N-fixers are responsible for a large fraction of biological soil-nitrogen inputs, which can increase nitrogen availability in forests in which N-fixers are locally abundant¹³. Symbioses with either N-fixers or ectomycorrhizal fungi often demand more plant photosynthate than does arbuscular mycorrhizal symbiosis^{12,14,15}. Because tree growth and reproduction are limited by access to inorganic, organic and atmospheric sources of nitrogen, the distribution of root symbioses is likely to reflect environmental conditions that maximize the cost:benefit ratio of symbiotic exchange as well as physiological constraints on the different symbionts.

One of the earliest efforts¹⁶ to understand the functional biogeography of plant root symbioses categorically classified biomes by their perceived dominant mycorrhizal type, and hypothesized that seasonal climates favour hosts that associate with ectomycorrhizal fungi (owing to the ability of these hosts to compete directly for organic nitrogen). By contrast, it has more recently been proposed that sensitivity to low temperatures has prevented N-fixers from dominating outside of the tropics, despite the potential for nitrogen fixation to alleviate nitrogen limitation in boreal forests^{15,17}. However, global-scale tests of

¹Department of Biology, Stanford University, Stanford, CA, USA. ²Department of Environmental Systems Science, ETH Zürich, Zürich, Switzerland. ³Department of Forestry and Natural Resources, Purdue University, West Lafayette, IN, USA. ⁴Research Center of Forest Management Engineering of State Forestry and Grassland Administration, Beijing Forestry University, Beijing, China.

⁵Department of Zoology, University of Oxford, Oxford, UK. ⁶Department of Forest Resources, University of Minnesota, St Paul, MN, USA. ⁷Hawkesbury Institute for the Environment, Western Sydney University, Penrith, New South Wales, Australia. ⁸Wageningen University and Research, Wageningen, The Netherlands. ⁹Department of Crop and Forest Sciences - Agrotecnio Center (UdL-Agrotecnio), Universitat de Lleida, Lleida, Spain. ¹⁰Forest Science and Technology Centre of Catalonia (CTFC), Solsona, Spain. ¹¹Food and Agriculture Organization of the United Nations, Rome, Italy. ¹²Cirad, UPR Forêts et Sociétés, University of Montpellier, Montpellier, France. ¹³Department of Forestry and Environment, National Polytechnic Institute (INP-HB), Yamoussoukro, Côte d'Ivoire. ¹⁴A list of participants and their affiliations appears in the online version of the paper. ¹⁵These authors contributed equally: B. S. Steidinger, T. W. Crowther, J. Liang.

*e-mail: tom.crowther@usys.ethz.ch; albeca.liang@gmail.com; kpeay@stanford.edu

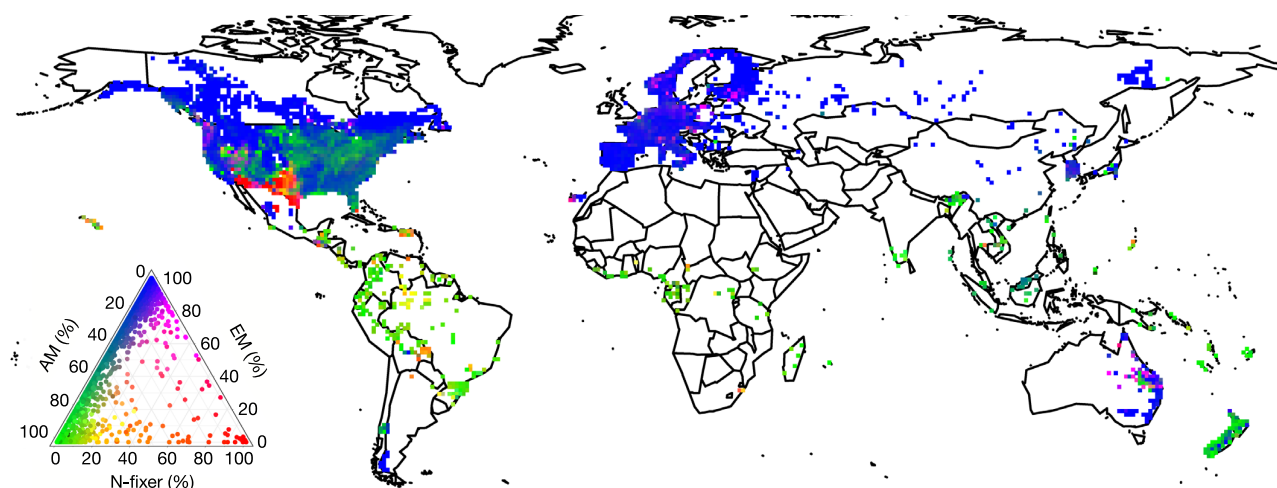


Fig. 1 | The global distribution of GFBi training data. The global map has $n = 2,768$ grid cells at a resolution of $1^\circ \times 1^\circ$ latitude and longitude. Cells are coloured in the red, green and blue spectrum according to

the percentage of total tree basal area occupied by N-fixer, arbuscular mycorrhizal (AM) and ectomycorrhizal (EM) tree symbiotic guilds, as indicated by the ternary plot.

these proposed biogeographical patterns and their climate drivers are lacking. To address this, we compiled a global ground-sourced survey database to reveal the numerical abundances of each type of symbiosis across the globe. Such a database is essential for identifying the potential mechanisms that underlie transitions in forest symbiotic state along climatic gradients^{18,19}.

We determined the abundance of tree symbioses using an extension of the plot-based Global Forest Biodiversity (GFB) database that we term the GFBi; this extended database contains over 1.1 million forest inventory plots of individual-based measurement records, from which we derive abundance information for entire tree communities (Fig. 1). Using published literature on the evolutionary histories of mycorrhizal and N-fixer symbioses, we assigned plant species from the GFBi to one of five root-associated symbiotic guilds: arbuscular mycorrhizal, ectomycorrhizal, ericoid mycorrhizal, N-fixer and weakly arbuscular or non-mycorrhizal. We then used the random-forest algorithm with K -fold cross-validation to determine the importance and influence of variables related to climate, soil chemistry, vegetation and topography on the relative abundance of each tree symbiotic guild (Fig. 2). Because decomposition is the dominant process by which soil nutrients become available to plants, we calculated annual and quarterly decomposition coefficients according to the Yasso07 model²⁰, which describes how temperature and precipitation gradients influence mass-loss rates of different chemical pools of leaf litter (with parameters fit using a previous global study of leaf decomposition) (Fig. 3, Supplementary Fig. 5). Finally, we projected our predictive models across the globe over the extent of global biomes that fell within the multivariate distribution of our model training data (Fig. 4, Supplementary Figs. 14, 15; see Methods for full description).

Our analysis shows that each one of the three most-numerically abundant guilds of tree symbiosis has a reliable environmental signature, with the four most-important predictors accounting for 81, 79 and 52% of the total variability in relative basal area for ectomycorrhizal, arbuscular mycorrhizal and N-fixer symbioses, respectively. Given the relative rarity of ericoid mycorrhizal and weakly arbuscular or non-mycorrhizal symbiotic states among trees, models for these symbioses lack strong predictive power—although the raw data do identify some local abundance hotspots for ericoid mycorrhizal symbiosis (Supplementary Fig. 1). As a result, we focus on the three major tree symbiotic states (ectomycorrhizal, arbuscular mycorrhizal and N-fixer). Despite the fact that data from North America and South America constitute 65% of the training data (at the 1° -by- 1° grid scale), our models accurately predict the proportional abundances of the three major symbioses across all major geographical regions (Supplementary Fig. 10). The high performance of our models—which is robust to

K -fold cross-validation and to rarefying samples such that all continents are represented with equal depth (Supplementary Figs. 11, 12)—suggests that regional variations in climate (including indirect effects on decomposition) and soil pH (for N-fixers) are the primary factors that influence the relative dominance of each guild at the global scale; geographical origin explained only approximately 2–5% of the variability in residual relative abundance (Supplementary Table 8, Supplementary Fig. 10).

Whereas a recent global analysis of root traits concluded that plant evolution has favoured a reduced dependence on mycorrhizal fungi²¹, we find that trees that associate with the relatively more carbon-demanding and recently derived ectomycorrhizal fungi^{12,14} represent the dominant tree symbiosis. By taking the average proportion of ectomycorrhizal trees, weighted by spatially explicit global predictions for tree stem density¹¹, we estimate that approximately 60% of tree stems on earth are ectomycorrhizal—despite the fact that only 2% of overall plant species associate with ectomycorrhizal fungi (versus nearly 80% that associate with arbuscular mycorrhizal fungi)⁷. Outside of the tropics, the estimate for the relative abundance of ectomycorrhizal symbiosis increases to approximately 80% of trees.

Turnover among the major symbiotic guilds results in a tri-modal latitudinal abundance gradient, in which the proportion of ectomycorrhizal trees increases (and the proportion of arbuscular mycorrhizal trees decreases) with distance from the equator and the upper quantiles of nitrogen-fixing trees reach a peak in abundance in the arid zone at around 30° N or S (Figs. 3a and 4). These trends are driven by abrupt transitional regions along continental climatic gradients (Fig. 2), which skew the distribution of symbioses among biomes (Fig. 3a) and drive strong patterns across geographical and topographic features that influence climate. Moving north or south from the equator, the first transitional zone separates warm (aseasonal) tropical broadleaf forests dominated by arbuscular mycorrhizal symbiosis (>75% median basal area versus 8% for ectomycorrhizal trees) from the rest of the world forest system, which is dominated by ectomycorrhizal symbiosis (Figs. 2a, b and 3a). The transition zone occurs across the globe at around 25° N and S, just beyond the dry tropical broadleaf forests (which have 25% of their basal area consisting of ectomycorrhizal trees) (Fig. 3a) in which average monthly temperature variation reaches 3 – 5° C (temperature seasonality) (Fig. 2a, b).

Moving further north or south, the second transitional climate zone separates regions in which decomposition coefficients during the warmest quarter of the year are less than two (Fig. 3b gives the associated temperature and precipitation ranges). In North America and China, this transition zone occurs around 50° N, and separates the mixed arbuscular mycorrhizal and ectomycorrhizal temperate forests

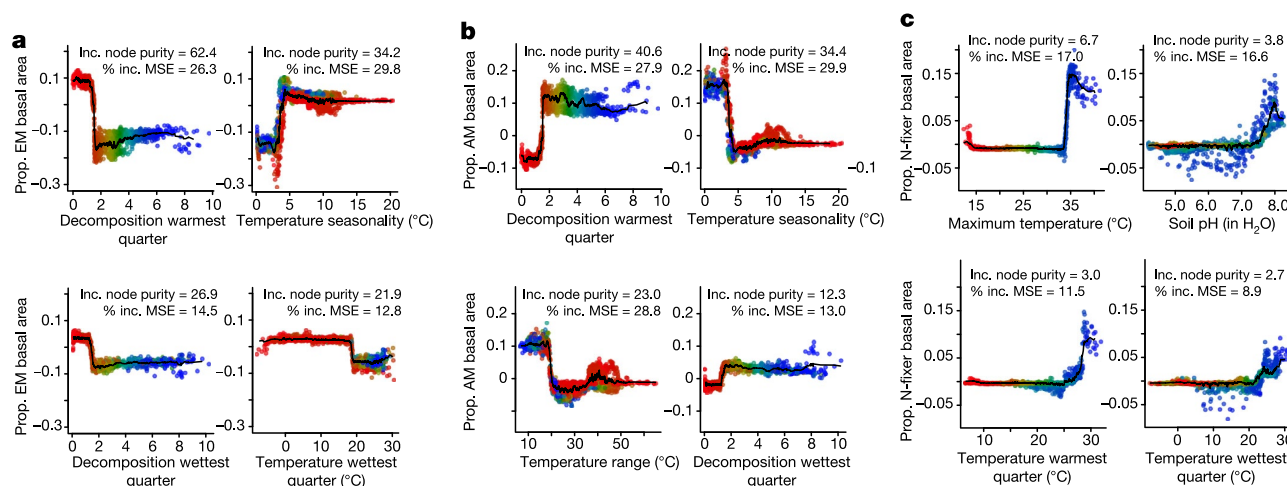


Fig. 2 | A small number of environmental variables predict the majority of global turnover in forest symbiotic status. **a–c**, Partial feature contributions of different environmental variables to forest symbiotic state. Each row plots the shape of the contribution of the four most-important predictors of the proportion of tree basal area that belongs to the ectomycorrhizal (**a**), arbuscular mycorrhizal (**b**) and N-fixer (**c**) symbiotic guilds ($n = 2,768$). Variables are listed in declining importance from left to right, as determined by the increase in node purity (inc. node purity), and with points coloured with a red to green to blue gradient

from their neighbouring ectomycorrhizal-dominated boreal forests (75 and 100% of their basal area, respectively, consisting of ectomycorrhizal trees) (Fig. 3a). This transitional decomposition zone is not present in western Europe, which has a temperature seasonality of $>5^{\circ}\text{C}$ but lacks sufficiently wet summers to accelerate decomposition coefficients beyond the values that are associated with mixed arbuscular mycorrhizal and ectomycorrhizal forests. The latitudinal transitions in symbiotic state observed among biomes are mirrored by within-biome transitions along elevation gradients. For example, in tropical Mexico decomposition coefficients of less than two during the warmest and wettest quarters of the year occur along the slopes of the Sierra Madre,

according to their position on the x axis of the most-important variable (left-most panels for each guild), allowing cross-visualization between predictors. Each panel lists two measures of variable importance; inc. node purity (used for sorting) and percentage increase in mean square error (% inc. MSE) (see Supplementary Information). The abundance of each type of symbiont transitions sharply along climatic gradients, which suggests that sites near the threshold are particularly vulnerable to switching their dominant symbiotic guild as climate changes. Warmest and wettest quarter, the warmest and wettest quarters of the year, respectively.

where a mixture of arbuscular-mycorrhizal and N-fixer woodlands in arid climates transition to ectomycorrhizal-dominated tropical coniferous forests (75% basal area) (Figs. 3a and 4a–c, Supplementary Figs. 16–18). The Southern Hemisphere—which lacks the landmass to support extensive boreal forests—experiences a similar latitudinal transition in decomposition rates along the ecotone that separates its tropical and temperate biomes, at around 28°S .

The abrupt transitions that we detected between forest symbiotic states along environmental gradients suggest that positive feedback effects may exist between climatic and biological controls of decomposition^{10,20}. In contrast to arbuscular mycorrhizal fungi, some

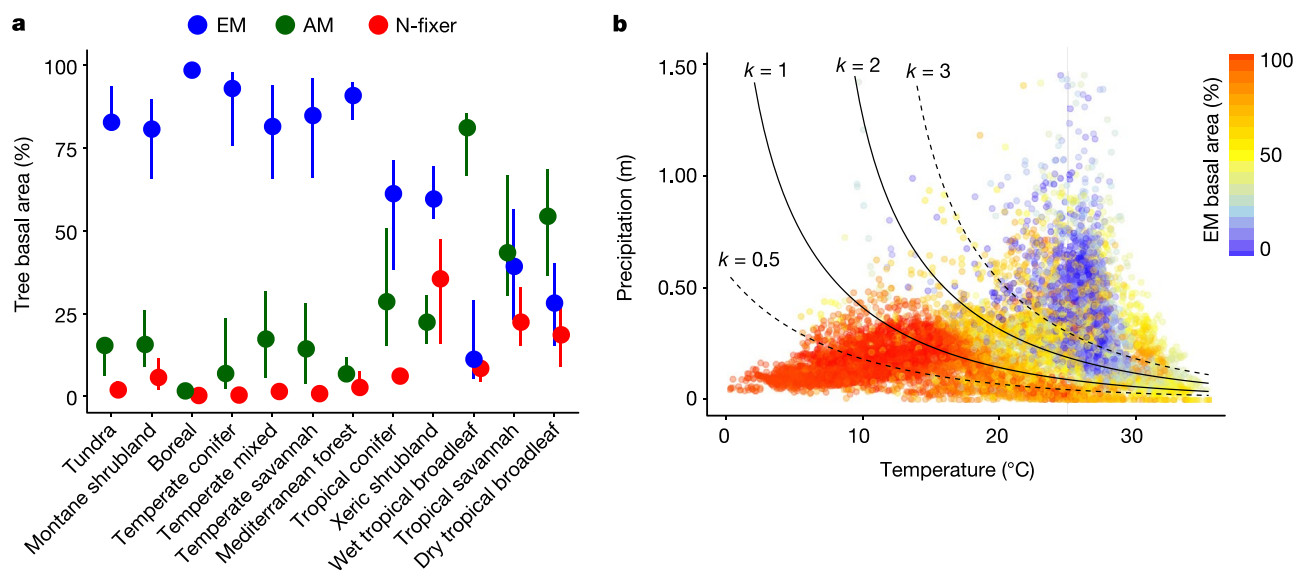


Fig. 3 | The distribution of forest symbiotic status across biomes is related to climatic controls over decomposition. **a**, Biome level summaries of the median ± 1 quartile of the predicted percentage of tree basal area per biome for ectomycorrhizal, arbuscular mycorrhizal and N-fixer symbiotic guilds ($n = 100$ random samples per biome). **b**, The dependency of decomposition coefficients (k , solid and dotted lines; in the region between the solid lines, the model transitions abruptly

between dominant symbiotic status) on temperature and precipitation during the warmest quarter with respect to predicted dominance of mycorrhizal symbiosis. The transition from arbuscular mycorrhizal forests to ectomycorrhizal forests between $k = 1$ and $k = 2$ is abrupt, which is consistent with positive feedback between climatic and biological controls of decomposition.

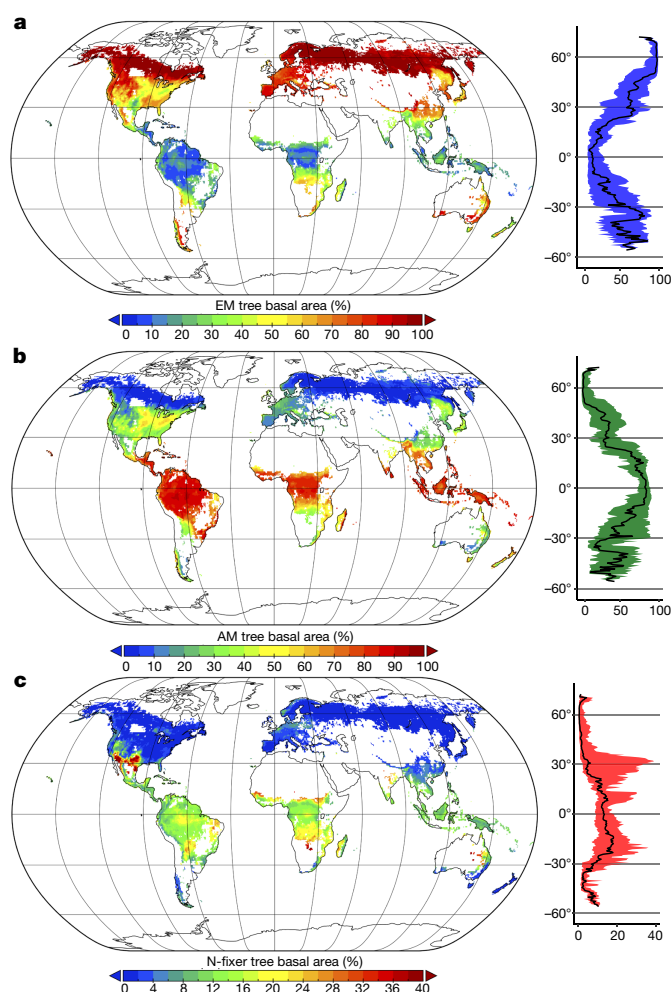


Fig. 4 | Global maps of predicted forest-tree symbiotic state. **a–c**, Maps (left) and latitudinal gradients (right; solid line indicates median; coloured ribbon spans the range between the 5% and 95% quantiles) of the percentage of tree basal area for ectomycorrhizal (**a**), arbuscular mycorrhizal (**b**) and N-fixer (**c**) symbiotic guilds. All projections are displayed on a 0.5°-by-0.5° latitude and longitude scale. $n = 28,454$ grid cells.

ectomycorrhizal fungi can use oxidative enzymes to mineralize organic nutrients from leaf litter and convert nutrients to plant-usable forms^{2,5}. Relative to arbuscular mycorrhizal trees, the leaf litter of ectomycorrhizal trees is also chemically more resistant to decomposition, and has higher C:N ratios and higher concentrations of decomposition-inhibiting secondary compounds¹⁰. Thus, ectomycorrhizal leaf litter can exacerbate climatic barriers to decomposition and promote conditions in which ectomycorrhizal fungi have nutrient-acquiring abilities that are superior to those of arbuscular mycorrhizal fungi^{5,10}. A recent game-theoretical model has shown that positive feedback effects between plants and soil nutrients can lead to local bistability in mycorrhizal symbiosis²². Such positive feedback effects are also known to cause abrupt ecosystem transitions along smooth environmental gradients between woodlands and grasses: trees suppress fires (which promotes seedling recruitment), whereas grass fuels fires that kill tree seedlings²³. The existence of abrupt transitions also suggests that forests in transitional regions along decomposition gradients should be susceptible to marked turnover in symbiotic state with future environmental changes²³.

To illustrate the sensitivity of global patterns of tree symbiosis to climate change, we use the relationships that we observed for current climates to project potential changes in the symbiotic status of forests in the future. Relative to our global predictions that use the most-recent

climate data, model predictions that use the projected climates for 2070 suggest that the abundance of ectomycorrhizal trees will decline by as much as 10% (using a relative concentration pathway of 8.5 W per m²) (Supplementary Fig. 24). Our models predict that the largest declines in ectomycorrhizal abundance will occur along the boreal–temperate ecotone, where small increases in climatic decomposition coefficients cause abrupt transitions to arbuscular mycorrhizal forests (Fig. 2a, b). Although our model does not estimate the time lag between climate change and forest community responses, the predicted decline in ectomycorrhizal trees corroborates the results of common garden transfer and simulated warming experiments, which have demonstrated that some important ectomycorrhizal hosts will decline at the boreal–temperate ecotone under altered climate conditions²⁴.

The change in dominant nutrient-exchange symbioses along climate gradients highlights the interconnection between atmospheric and soil compartments of the biosphere. The transition from arbuscular mycorrhizal to ectomycorrhizal dominance corresponds with a shift from phosphorus to nitrogen limitation of plant growth with increasing latitude^{25,26}. Including published global projections of total soil nitrogen or phosphorus, microbial nitrogen or soil phosphorus fractions (labile, occluded, organic and apatite) did not increase the amount of variation explained by the model, or alter the variables identified as most important; we therefore dropped these projections from our analysis. However, our finding that climatic controls of decomposition are the best predictors of dominant mycorrhizal associations provides a mechanistic link between symbiont physiology and climatic controls on the release of soil nutrients from leaf litter. These findings are consistent with Read's hypothesis¹⁶ that slow decomposition at high latitudes favours ectomycorrhizal fungi owing to their increased capacity to liberate organic nutrients². Thus, although more experiments are necessary to understand the specific mechanism by which nutrient competition favours the dominance of arbuscular mycorrhizal or ectomycorrhizal symbioses¹⁸, we propose that the latitudinal and elevational transitions from arbuscular-mycorrhizal-dominated to ectomycorrhizal-dominated forests be named 'Read's rule'.

Our analyses focus on prediction at large spatial scales that are appropriate to the available data, but our findings with respect to Read's rule also provide insight into how soil factors structure the fine-scale distributions of tree symbioses within our grid cells. For example, at a coarse scale, we find that ectomycorrhizal trees are relatively rare in many wet tropical forests; however, individual tropical sites in our raw data span the full range from 0 to 100% basal area dominated by ectomycorrhizal trees. In much of the wet tropics, these ectomycorrhizal-dominated sites exist as outliers within a matrix of predominantly arbuscular mycorrhizal trees. In an apparent exception that proves Read's rule, in aseasonal, warm neotropical climates—which accelerate leaf decomposition and promote the regional dominance of arbuscular mycorrhizal symbiosis (Fig. 3)—ectomycorrhizal-dominated tree stands can develop in sites in which poor soils and recalcitrant litter slow the rates of decomposition and nitrogen mineralization^{18,27}. Landscape-scale variation in the relative abundance of symbiotic states also changes along climate gradients: variability is highest in xeric and temperate biomes (Supplementary Figs. 3, 4), which suggests that the potential of local nutrient variability to favour particular symbioses is contingent on climate.

Whereas ectomycorrhizal trees are associated with ecosystems in which plant growth is thought to be primarily nitrogen-limited, N-fixer trees are not. Our results highlight the global extent of the apparent 'nitrogen cycling paradox' in which some metrics suggest that nitrogen limitation is greater in the temperate zone^{25,26} and yet nitrogen-fixing trees are relatively more common in the tropics^{15,28} (Fig. 3a). We find that N-fixers—which we estimate represent 7% of all trees—dominate forests with annual maximum temperatures >35 °C and alkaline soils, particularly in North America and Africa (Fig. 2c). N-fixers have the highest relative abundance in xeric shrublands (24%), tropical savannahs (21%) and dry broadleaf forest biomes (20%), but are nearly absent from boreal forests (<1%) (Figs. 3a and 4). The decline in N-fixer tree

abundance with increasing latitude that we observed is also associated with a previously documented latitudinal shift in the identity of nitrogen-fixing microorganisms, from facultative rhizobial N-fixers in tropical forests to obligate actinorhizal N-fixers in temperate forests²⁸. Our data are not capable of fully disentangling the several hypotheses that have previously been proposed to reconcile the nitrogen cycling paradox¹⁵. However, our results are consistent with the model prediction¹⁷ and regional empirical evidence^{19,29,30} that nitrogen-fixing trees are particularly important in arid biomes. Based primarily on the observed positive nonlinear association of the relative abundance of N-fixers with the mean temperature of the hottest month (Fig. 2c), our models predict a twofold increase in relative abundance of N-fixers when transitioning from humid to dry tropical forest biomes (Fig. 3a).

Although soil microorganisms are a dominant component of forests in terms of both diversity and ecosystem functioning^{5,6,10}, identifying global-scale microbial biogeographical patterns remains an ongoing research priority. Our analyses confirm that Read's rule—which is one of the first proposed biogeographical rules specific to microbial symbioses—successfully describes global transitions between mycorrhizal guilds. More generally, climate driven turnover among the major symbioses between plants and microorganisms represents a fundamental biological pattern in the Earth system, as forests transition from low-latitude arbuscular mycorrhizal through N-fixer to high-latitude ectomycorrhizal ecosystems. The predictions of our model (available in the Supplementary Data as global raster layers) can now be used to represent these critical ecosystem variations in global biogeochemical models that are used to predict climate–biogeochemical feedback effects within and between trees, soils and the atmosphere. Additionally, the raster layer that contains the proportion of nitrogen-fixing trees can be used to map potential symbiotic nitrogen fixation, which links atmospheric pools of carbon and nitrogen. Future work can extend our findings to incorporate multiple plant growth forms and non-forested biomes (in which similar patterns are likely to exist) to generate a complete global perspective. Our predictive maps leverage a comprehensive global forest dataset to generate a quantitative global map of forest tree symbioses, and demonstrate how nutritional mutualisms are coupled with the global distribution of plant communities.

Online content

Any methods, additional references, Nature Research reporting summaries, source data, statements of data availability and associated accession codes are available at <https://doi.org/10.1038/s41586-019-1128-0>.

Received: 27 April 2018; Accepted: 21 March 2019;

Published online 15 May 2019.

1. Batterman, S. A. et al. Key role of symbiotic dinitrogen fixation in tropical forest secondary succession. *Nature* **502**, 224–227 (2013).
2. Shah, F. et al. Ectomycorrhizal fungi decompose soil organic matter using oxidative mechanisms adapted from saprotrophic ancestors. *New Phytol.* **209**, 1705–1719 (2016).
3. Averill, C., Turner, B. L. & Finzi, A. C. Mycorrhiza-mediated competition between plants and decomposers drives soil carbon storage. *Nature* **505**, 543–545 (2014).
4. Clemmensen, K. E. et al. Roots and associated fungi drive long-term carbon sequestration in boreal forest. *Science* **339**, 1615–1618 (2013).
5. Cheeke, T. E. et al. Dominant mycorrhizal association of trees alters carbon and nutrient cycling by selecting for microbial groups with distinct enzyme function. *New Phytol.* **214**, 432–442 (2017).
6. Terrer, C., Vicca, S., Hungate, B. A., Phillips, R. P. & Prentice, I. C. Mycorrhizal association as a primary control of the CO₂ fertilization effect. *Science* **353**, 72–74 (2016).
7. Brundrett, M. C. & Tedersoo, L. Evolutionary history of mycorrhizal symbioses and global host plant diversity. *New Phytol.* **220**, 1108–1115 (2018).
8. Averill, C. & Hawkes, C. V. Ectomycorrhizal fungi slow soil carbon cycling. *Ecol. Lett.* **19**, 937–947 (2016).
9. Bennett, J. A. et al. Plant–soil feedbacks and mycorrhizal type influence temperate forest population dynamics. *Science* **355**, 181–184 (2017).
10. Phillips, R. P., Brzostek, E. & Midgley, M. G. The mycorrhizal-associated nutrient economy: a new framework for predicting carbon–nutrient couplings in temperate forests. *New Phytol.* **199**, 41–51 (2013).

11. Crowther, T. W. et al. Mapping tree density at a global scale. *Nature* **525**, 201–205 (2015).
12. van der Heijden, M. G., Martin, F. M., Selosse, M. A. & Sanders, I. R. Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytol.* **205**, 1406–1423 (2015).
13. Binkley, D., Sollins, P., Bell, R., Sachs, D. & Myrold, D. Biogeochemistry of adjacent conifer and alder-conifer stands. *Ecology* **73**, 2022–2033 (1992).
14. Leake, J. et al. Networks of power and influence: the role of mycorrhizal mycelium in controlling plant communities and agroecosystem functioning. *Can. J. Bot.* **82**, 1016–1045 (2004).
15. Hedin, L. O., Brookshire, E. N. J., Menge, D. N. L. & Barron, A. R. The nitrogen paradox in tropical rainforest ecosystems. *Annu. Rev. Ecol. Evol. Syst.* **40**, 613–635 (2009).
16. Read, D. J. Mycorrhizas in ecosystems. *Experientia* **47**, 376–391 (1991).
17. Houlton, B. Z., Wang, Y.-P., Vitousek, P. M. & Field, C. B. A unifying framework for dinitrogen fixation in the terrestrial biosphere. *Nature* **454**, 327–330 (2008).
18. Peay, K. G. The mutualistic niche: mycorrhizal symbiosis and community dynamics. *Annu. Rev. Ecol. Evol. Syst.* **47**, 143–164 (2016).
19. Pellegrini, A. F. A., Staver, A. C., Hedin, L. O., Charles-Dominique, T. & Tourgee, A. Aridity, not fire, favors nitrogen-fixing plants across tropical savanna and forest biomes. *Ecology* **97**, 2177–2183 (2016).
20. Tuomi, M. et al. Leaf litter decomposition—estimates of global variability based on Yasso07 model. *Ecol. Modell.* **220**, 3362–3371 (2009).
21. Ma, Z. et al. Evolutionary history resolves global organization of root functional traits. *Nature* **555**, 94–97 (2018).
22. Lu, M. & Hedin, L. O. Global plant–symbiont organization and emergence of biogeochemical cycles resolved by evolution-based trait modelling. *Nat. Ecol. Evol.* **3**, 239–250 (2019).
23. Scheffer, M., Carpenter, S., Foley, J. A., Folke, C. & Walker, B. Catastrophic shifts in ecosystems. *Nature* **413**, 591–596 (2001).
24. Reich, P. B. et al. Geographic range predicts photosynthetic and growth response to warming in co-occurring tree species. *Nat. Clim. Change* **5**, 148 (2015).
25. McGroddy, M. E., Daufresne, T. & Hedin, L. O. Scaling of C:N:P stoichiometry in forests worldwide: implications of terrestrial redfield-type ratios. *Ecology* **85**, 2390–2401 (2004).
26. Reich, P. B. & Oleksyn, J. Global patterns of plant leaf N and P in relation to temperature and latitude. *Proc. Natl Acad. Sci. USA* **101**, 11001–11006 (2004).
27. Corrales, A., Mangan, S. A., Turner, B. L. & Dalling, J. W. An ectomycorrhizal nitrogen economy facilitates monodominance in a neotropical forest. *Ecol. Lett.* **19**, 383–392 (2016).
28. Menge, D. N., Lichstein, J. W. & Angeles-Pérez, G. Nitrogen fixation strategies can explain the latitudinal shift in nitrogen-fixing tree abundance. *Ecology* **95**, 2236–2245 (2014).
29. Liao, W., Menge, D. N. L., Lichstein, J. W. & Angeles-Pérez, G. Global climate change will increase the abundance of symbiotic nitrogen-fixing trees in much of North America. *Glob. Change Biol.* **23**, 4777–4787 (2017).
30. Gei, M. et al. Legume abundance along successional and rainfall gradients in neotropical forests. *Nat. Ecol. Evol.* **2**, 1104–1111 (2018).

Acknowledgements This work was made possible by the Global Forest Biodiversity Database, which represents the work of over 200 independent investigators and their public and private funding agencies (see Supplementary Acknowledgements).

Reviewer information *Nature* thanks Martin Bidartondo, David Bohan and the other anonymous reviewer(s) for their contribution to the peer review of this work.

Author contributions K.G.P. and T.W.C. conceived the study; T.W.C., J.L., P.B.R., G.N., S.d.-M., M.Z., N.P., B.H., X.Z. and C.Z. conceived and organized the GFB database; K.G.P., B.S.S., G.D.A.W. and M.E.V.N. compiled the symbiosis database; B.S.S. carried out the primary data analysis; M.E.V.N. and D.R. contributed to data compilation and analysis; B.S.S., T.W.C., M.E.V.N. and K.G.P. wrote the initial manuscript; B.S.S., T.W.C., J.L., M.E.V.N., G.D.A.W., P.B.R., G.N., S.d.-M., M.Z., N.P., B.H., X.Z., C.Z. and K.G.P. made substantial revisions to all versions of the manuscript; all other named authors provided forest inventory data and commented on the manuscript.

Competing interests The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41586-019-1128-0>.

Reprints and permissions information is available at <http://www.nature.com/reprints>.

Correspondence and requests for materials should be addressed to T.W.C., J.L. or K.G.P.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© The Author(s), under exclusive licence to Springer Nature Limited 2019

GFBI consortium

Meinrad Abegg¹⁶, C. Yves Adou Yao¹⁷, Giorgio Alberti^{18,19}, Angelica Almeyda Zambrano²⁰, Esteban Alvarez-Davila²¹, Patricia Alvarez-Loayza²², Luciana F. Alves²³, Christian Ammer²⁴, Clara Antón-Fernández²⁵, Alejandro Araujo-Murakami²⁶, Luzmila Arroyo²⁶, Valerio Avitabile²⁷, Gerardo Aymard²⁸, Timothy Baker²⁹, Radomir Balazy³⁰, Olaf Banki³¹, Jorcelly Barroso³², Meredith Bastian³³, Jean-Francois Bastin², Luca Biragazzi¹¹, Philippe Birnbaum¹⁶², Robert Bitariho³⁴, Pascal Boeckx³⁵, Frans Bongers, Olivier Bouriaud³⁶, Pedro H. S. Brancalion³⁷, Susanne Brandl³⁸, Francis Q. Brearley³⁹, Roel Brienens²⁹, Eben Broadbent¹⁷⁵, Helge Bruehlheide^{40,41}, Filippo Bussotti⁴², Roberto Cazzolla Gatti⁴³, Ricardo Cesar³⁷, Goran Cesljar⁴⁴, Robin Chazdon^{45,46}, Han Y. H. Chen^{47,48}, Chelsea Chisholm⁴⁹, Emil Cienciala^{50,51}, Connie J. Clark⁵², David Clark⁵³, Gabriel Colletta¹⁷⁴, Richard Condit⁵⁵, David Coomes⁵⁶, Fernando Cornejo Valverde⁵⁷, Jose J. Corral-Rivas⁵⁸, Philip Crim^{59,60}, Jonathan Cumming⁶⁰, Selvadurai Dayanandan⁶¹, André L. de Gasper⁶², Mathieu Decuyper⁶³, Gérard Delroire⁶⁴, Ben DeVries⁶⁴, Ilija Djordjevic⁶⁵, Amaral Iêda⁶⁶, Aurélie Dourdain⁶³, Nestor Laurier Engone Obiang⁶⁷, Brian Enquist^{68,69}, Teresa Eyre⁷⁰, Adandé Belarmain Fandohan⁷¹, Tom M. Fayle⁷², Ted R. Feldpausch⁷³, Leena Finér⁷⁴, Markus Fischer⁷⁵, Christine Fletcher⁷⁶, Jonas Friedman⁷⁷, Lorenzo Frizzera⁷⁸, Javier G. P. Gamarra¹¹, Damiano Gianelle⁷⁸, Henry B. Glick⁷⁹, David Harris⁸⁰, Andrew Hector⁸¹, Andreas Hemp⁸², Geerten Hengeveld⁸, John Herbohn⁴⁶, Martin Herold⁸, Annika Hillers^{83,178}, Euridice N. Honorio Coronado⁸⁴, Markus Huber¹⁶, Cang Hui^{85,86}, Hyunkook Cho⁸⁷, Thomas Ibanez⁸⁸, Ilbin Jung⁸⁷, Nobuo Imai⁸⁹, Andrzej M. Jagodzinski^{90,91}, Bogdan Jaroszewicz⁹², Vivian Johannsen⁹³, Carlos A. Joly⁵⁴, Tommaso Jucker⁹⁴, Viktor Karminov⁹⁵, Kuswata Kartawinata²², Elizabeth Kearsley⁹⁶, David Kenfack⁹⁷, Deborah Kennard⁹⁸, Sebastian Kepfer-Rojas⁹³, Gunnar Keppel⁹⁹, Mohammed Latif Khan¹⁰⁰, Timothy Killeen²⁶, Hyun Seok Kim^{101,102,103,104}, Kanehiro Kitayama¹⁰⁵, Michael Köhl¹⁰⁶, Henn Korjus¹⁰⁷, Florian Kraxner¹⁰⁸, Diana Laarmann¹⁰⁷, Mait Lang¹⁰⁷, Simon Lewis^{29,109}, Huicui Lu¹¹⁰, Natalia Lukina¹¹¹, Brian Maitner⁶⁸, Yadvinder Malhi¹¹², Eric Marcon¹¹³, Beatriz Schwantes Marimon¹¹⁴, Ben Hur Marimon-Junior¹¹⁴, Andrew Robert Marshall^{46,115,176}, Emanuel Martin¹¹⁶, Olga Martynenko⁹⁵, Jorge A. Meave¹¹⁷, Omar Melo-Cruz¹¹⁸, Casimiro Mendoza¹¹⁹, Cory Merow⁴⁵, Abel Monteagudo Mendoza^{120,121}, Vanessa Moreno³⁷, Sharif A. Mukul^{46,122}, Philip Mundhenk¹⁰⁶, Maria G. Nava-Miranda¹²³, David Neill¹²⁴, Victor Neldner⁷⁰, Radovan Nevenic⁶⁵, Michael Ngugi⁷⁰, Pascal Niklaus¹²⁵, Jacek Oleksyn⁹⁰, Petr Ontikov⁹⁵, Edgar Ortiz-Malavasi¹²⁶, Yude Pan¹²⁷, Alain Paquette¹²⁸, Alexander Parada-Gutiérrez²⁶, Elena Parfenova¹²⁹, Minjee Park¹⁰¹, Marc Parren¹³⁰, Narayanawamy Parthasarathy¹³¹, Pablo L. Peri¹³², Sebastian Pfautsch¹³³, Oliver Phillips²⁹, Maria Teresa Piedade¹³⁴, Daniel Piotto¹³⁵, Nigel C. A. Pitman²², Irina Polo¹³⁶, Lourens Poorter⁸, Axel Dalberg Poulsen⁸⁰, John R. Poulsen⁵², Hans Pretzsch¹³⁷, Freddy Ramirez Arevalo¹³⁸, Zorayda Restrepo-Correa¹³⁹, Mirco Rodighiero^{78,177}, Samir Rolim¹³⁵, Anand Roopsind¹⁴⁰, Francesco Rovero^{141,142}, Ervan Rutishauser⁵⁵, Purabi Saikia¹⁴³, Philippe Saner¹²⁵, Peter Schall²⁴, Mart-Jan Schelhaas⁸, Dmitry Schepaschenko¹⁰⁸, Michael Scherer-Lorenzen¹⁴⁴, Bernhard Schmid¹²⁵, Jochen Schöngart¹³⁴, Eric Searle⁴⁷, Vladimir Seben¹⁴⁵, Josep M. Serra-Diaz^{146,147}, Christian Salas-Eljatib^{179,180}, Douglas Sheil¹⁸¹, Anatoly Shvidenko¹⁰⁸, Javier Silva-Espejo¹⁴⁸, Marcos Silveira¹⁴⁹, James Singh¹⁵⁰, Plinio Sist¹², Ferry Slik¹⁵¹, Bonaventure Sonké¹⁵², Alexandre F. Souza¹⁵³, Krzysztof Stereńczak³⁰, Jens-Christian Svenning^{147,154}, Miroslav Svoboda¹⁵⁵, Natalia Targhetta¹³⁴, Nadja Tchebakova¹²⁹, Hans ter Steege^{31,156}, Raquel Thomas¹⁵⁷, Elena Tikhonova¹¹¹, Peter Umunay⁷⁹, Vladimir Usoltsev¹⁵⁸, Fernando Valladares¹⁵⁹, Fons van der Plas¹⁶⁰, Tran Van Do¹⁶¹, Rodolfo Vasquez Martinez¹²⁰, Hans Verbeecq⁹⁶, Helder Viana^{163,164}, Simone Vieira¹⁶⁵, Klaus von Gadow¹⁶⁶, Hua-Feng Wang¹⁶⁷, James Watson¹⁶⁸, Bertil Westerlund⁷⁷, Susan Wiser¹⁶⁹, Florian Wittmann¹⁷⁰, Virginia Wortel¹⁷¹, Roderick Zagt¹⁷², Tomasz Zawila-Niedzwiecki¹⁷³, Zhi-Xin Zhu¹⁶⁷ & Irie Casimir Zo-Bi¹³

¹⁶Swiss Federal Institute for Forest, Snow and Landscape Research, WSL, Birmensdorf, Switzerland. ¹⁷UFR Biosciences, University Félix Houphouët-Boigny, Abidjan, Côte d'Ivoire. ¹⁸Department of Agricultural, Food, Environmental and Animal Sciences, University of Udine, Udine, Italy. ¹⁹Institute of Biometeorology, National Research Council (CNR-IBIMET), Florence, Italy. ²⁰Spatial Ecology and Conservation Laboratory, Department of Tourism, Recreation and Sport Management, University of Florida, Gainesville, FL, USA. ²¹Fundación ConVida, Universidad Nacional Abierta y a Distancia, UNAD, Medellín, Colombia. ²²Field Museum of Natural History, Chicago, IL, USA. ²³Center for Tropical Research, Institute of the Environment and Sustainability, UCLA, Los Angeles, CA, USA. ²⁴Silviculture and Forest Ecology of the Temperate Zones, University of Göttingen, Göttingen, Germany. ²⁵Division of Forest and Forest Resources, Norwegian Institute of Bioeconomy Research (NIBIO), Ås, Norway. ²⁶Museo de Historia Natural Noel Kempff Mercado, Universidad Autónoma Gabriel René Moreno, Santa Cruz de la Sierra, Bolivia. ²⁷European Commission, Joint Research Center, Ispra, Italy. ²⁸UNELLEZ-Guanare, Programa de Ciencias del Agro y el Mar, Herbario Universitario (PORT), Portuguesa, Venezuela. ²⁹School of Geography, University of Leeds, Leeds, UK. ³⁰Department of Geomatics, Forest Research Institute, Raszyn, Poland. ³¹Naturalis Biodiversity Center, Leiden, The Netherlands. ³²Centro Multidisciplinar, Universidade Federal do Acre, Rio Branco, Brazil.

³³Smithsonian's National Zoo and Conservation Biology Institute, Washington, DC, USA.

³⁴Institute of Tropical Forest Conservation, Mbarara University of Sciences and Technology, Mbarara, Uganda. ³⁵Isotope Bioscience Laboratory - ISOFYS, Ghent University, Ghent, Belgium. ³⁶Integrated Center for Research, Development and Innovation in Advanced Materials, Nanotechnologies, and Distributed Systems for Fabrication and Control (MANSID), Stefan cel Mare University of Suceava, Suceava, Romania. ³⁷Department of Forest Sciences, Luiz de Queiroz College of Agriculture, University of São Paulo, Piracicaba, Brazil. ³⁸Bavarian State Institute of Forestry, Freising, Germany. ³⁹Manchester Metropolitan University, Manchester, UK. ⁴⁰Institute of Biology, Geobotany and Botanical Garden, Martin Luther University Halle-Wittenberg, Halle-Wittenberg, Germany. ⁴¹German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig, Leipzig, Germany. ⁴²Department of Agriculture, Food, Environment and Forest (DAGRI), University of Firenze, Florence, Italy. ⁴³Biological Institute, Tomsk State University, Tomsk, Russia. ⁴⁴Department of Spatial Regulation, GIS and Forest Policy, Institute of Forestry, Belgrade, Serbia. ⁴⁵Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, CT, USA. ⁴⁶Tropical Forests and People Research Centre, University of the Sunshine Coast, Maroochydore, Queensland, Australia. ⁴⁷Faculty of Natural Resources Management, Lakehead University, Thunder Bay, Ontario, Canada. ⁴⁸Key Laboratory for Humid Subtropical Eco-Geographical Processes of the Ministry of Education, Fujian Normal University, Fuzhou, China. ⁴⁹Institute of Integrative Biology, ETH Zürich, Zurich, Switzerland. ⁵⁰IFER - Institute of Forest Ecosystem Research, Jilove u Prahy, Czech Republic. ⁵¹Global Change Research Institute CAS, Brno, Czech Republic. ⁵²Nicholas School of the Environment, Duke University, Durham, NC, USA. ⁵³Department of Biology, University of Missouri-St Louis, St Louis, MO, USA. ⁵⁴Department of Plant Biology, Institute of Biology, University of Campinas, UNICAMP, Campinas, Brazil. ⁵⁵Smithsonian Tropical Research Institute, Balboa, Panama. ⁵⁶Department of Plant Sciences, University of Cambridge, Cambridge, UK. ⁵⁷Andes to Amazon Biodiversity Program, Madre de Dios, Peru. ⁵⁸Facultad de Ciencias Forestales, Universidad Juárez del Estado de Durango, Durango, Mexico. ⁵⁹Department of Physical and Biological Sciences, The College of Saint Rose, Albany, NY, USA. ⁶⁰Department of Biology, West Virginia University, Morgantown, WV, USA. ⁶¹Biology Department, Concordia University, Montreal, Quebec, Canada. ⁶²Natural Science Department, Universidade Regional de Blumenau, Blumenau, Brazil. ⁶³Cirad, UMR EcoFoG, Kourou, French Guiana. ⁶⁴Department of Geographical Sciences, University of Maryland, College Park, MD, USA. ⁶⁵Institute of Forestry, Belgrade, Serbia. ⁶⁶National Institute of Amazonian Research, Manaus, Brazil. ⁶⁷IRET, Herbar National du Gabon (CENAREST), Libreville, Gabon. ⁶⁸Department of Ecology and Evolutionary Biology, University of Arizona, Tucson, AZ, USA. ⁶⁹The Santa Fe Institute, Santa Fe, NM, USA. ⁷⁰Department of Environment and Science, Queensland Herbarium, Toowoomba, Queensland, Australia. ⁷¹Ecole de Foresterie et Ingénierie du Bois, Université Nationale d'Agriculture, Ketou, Benin. ⁷²Biology Centre of the Czech Academy of Sciences, Institute of Entomology, Ceske Budejovice, Czech Republic. ⁷³Geography, College of Life and Environmental Sciences, University of Exeter, Exeter, UK. ⁷⁴Natural Resources Institute Finland (Luke), Joensuu, Finland. ⁷⁵Institute of Plant Sciences, University of Bern, Bern, Switzerland. ⁷⁶Forest Research Institute Malaysia, Kuala Lumpur, Malaysia. ⁷⁷Department of Forest Resource Management, Swedish University of Agricultural Sciences SLU, Umeå, Sweden. ⁷⁸Department of Sustainable Agro-Ecosystems and Bioresources, Research and Innovation Center, Fondazione Edmund Mach, San Michele all'Adige, Italy. ⁷⁹School of Forestry and Environmental Studies, Yale University, New Haven, CT, USA. ⁸⁰Royal Botanic Garden Edinburgh, Edinburgh, UK. ⁸¹Department of Plant Sciences, University of Oxford, Oxford, UK. ⁸²Department of Plant Systematics, University of Bayreuth, Bayreuth, Germany. ⁸³Centre for Conservation Science, The Royal Society for the Protection of Birds, Sandy, UK. ⁸⁴Instituto de Investigaciones de la Amazonia Peruana, Iquitos, Peru. ⁸⁵Centre for Invasion Biology, Department of Mathematical Sciences, Stellenbosch University, Stellenbosch, South Africa. ⁸⁶Theoretical Ecology Unit, African Institute for Mathematical Sciences, Cape Town, South Africa. ⁸⁷Division of Forest Resources Information, Korea Forest Promotion Institute, Seoul, South Korea. ⁸⁸Institut Agronomique néo-Calédonien (IAC), Equipe Sol & Végétation (SolVeg), Nouméa, New Caledonia. ⁸⁹Department of Forest Science, Tokyo University of Agriculture, Tokyo, Japan. ⁹⁰Institute of Dendrology, Polish Academy of Sciences, Kórnik, Poland. ⁹¹Poznań University of Life Sciences, Department of Game Management and Forest Protection, Poznań, Poland. ⁹²Faculty of Biology, Białowieża Geobotanical Station, University of Warsaw, Białowieża, Poland. ⁹³Department of Geosciences and Natural Resource Management, University of Copenhagen, Copenhagen, Denmark. ⁹⁴Centre for Environment and Life Sciences, CSIRO Land and Water, Floreat, Western Australia, Australia. ⁹⁵Forestry Faculty, Bauman Moscow State Technical University, Mytitschi, Russia. ⁹⁶CAVElab - Computational and Applied Vegetation Ecology, Department of Environment, Ghent University, Ghent, Belgium. ⁹⁷CTFS-ForestGEO, Smithsonian Tropical Research Institute, Balboa, Panama. ⁹⁸Department of Physical and Environmental Sciences, Colorado Mesa University, Grand Junction, CO, USA. ⁹⁹School of Natural and Built Environments and Future Industries Institute, University of South Australia, Adelaide, South Australia, Australia. ¹⁰⁰Department of Botany, Dr Harisingh Gour Central University, Sagar, India. ¹⁰¹Department of Forest Sciences, Seoul National University, Seoul, South Korea. ¹⁰²Interdisciplinary Program in Agricultural and Forest Meteorology, Seoul National University, Seoul, South Korea. ¹⁰³National Center for Agro Meteorology, Seoul, South Korea. ¹⁰⁴Research Institute for Agriculture and Life Sciences, Seoul National University, Seoul, South Korea. ¹⁰⁵Graduate School of Agriculture, Kyoto University, Kyoto, Japan. ¹⁰⁶Institute for World Forestry, University of Hamburg, Hamburg, Germany. ¹⁰⁷Institute of Forestry and Rural Engineering, Estonian University of Life Sciences, Tartu, Estonia. ¹⁰⁸Ecosystems Services and Management, International Institute for Applied Systems Analysis, Laxenburg, Austria. ¹⁰⁹Department of Geography, University College London, London, UK. ¹¹⁰Faculty of Forestry, Qingdao Agricultural University, Qingdao, China. ¹¹¹Center for Forest Ecology and Productivity, Russian Academy of Sciences, Moscow, Russia. ¹¹²School of Geography, University of Oxford, Oxford, UK. ¹¹³UMR EcoFoG, AgroParisTech, Kourou, France. ¹¹⁴Departamento de Ciências Biológicas, Universidade do Estado de Mato Grosso, Nova Xavantina, Brazil. ¹¹⁵Department of Environment & Geography, University of York, York, UK. ¹¹⁶Department of Wildlife Management, College of African Wildlife Management, Mweka, Tanzania. ¹¹⁷Departamento de Ecología y Recursos Naturales, Facultad de Ciencias, Universidad Nacional Autónoma de México, Mexico

- City, Mexico. ¹¹⁸Universidad del Tolima, Ibagué, Colombia. ¹¹⁹Colegio de Profesionales Forestales de Cochabamba, Cochabamba, Bolivia. ¹²⁰Jardín Botánico de Missouri, Oxapampa, Peru. ¹²¹Universidad Nacional de San Antonio Abad del Cusco, Cusco, Peru. ¹²²Department of Environmental Management, School of Environmental Science and Management, Independent University Bangladesh, Dhaka, Bangladesh. ¹²³Instituto de Silvicultura e Industria de la Madera, Universidad Juárez del Estado de Durango, Durango, Mexico. ¹²⁴Universidad Estatal Amazónica, Puyo, Pastaza, Ecuador. ¹²⁵Department of Evolutionary Biology and Environmental Studies, University of Zürich, Zürich, Switzerland. ¹²⁶Forestry School, Tecnológico de Costa Rica TEC, Cartago, Costa Rica. ¹²⁷Climate, Fire, and Carbon Cycle Sciences, USDA Forest Service, Durham, NC, USA. ¹²⁸Centre for Forest Research, Université du Québec à Montréal, Montréal, Quebec, Canada. ¹²⁹V. N. Sukachev Institute of Forest, FRC KSC, Siberian Branch of the Russian Academy of Sciences, Krasnoyarsk, Russia. ¹³⁰Department of Forestry, World Research Institute, Washington, DC, USA. ¹³¹Department of Ecology and Environmental Sciences, Pondicherry University, Puducherry, India. ¹³²Instituto Nacional de Tecnología Agropecuaria (INTA), Universidad Nacional de la Patagonia Austral (UNPA), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Rio Gallegos, Argentina. ¹³³School of Social Sciences and Psychology (Urban Studies), Western Sydney University, Penrith, New South Wales, Australia. ¹³⁴Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil. ¹³⁵Laboratório de Dendrologia e Silvicultura Tropical, Centro de Formação em Ciências Agroflorestais, Universidade Federal do Sul da Bahia, Itabuna, Brazil. ¹³⁶Jardín Botánico de Medellín, Medellín, Colombia. ¹³⁷Chair for Forest Growth and Yield Science, TUM School for Life Sciences, Technical University of Munich, Munich, Germany. ¹³⁸Universidad Nacional de la Amazonia Peruana, Iquitos, Peru. ¹³⁹Servicios Ecosistémicos y Cambio Climático (SECC), Fundación Con Vida & Corporación COL-TREE, Medellín, Colombia. ¹⁴⁰Department of Biological Sciences, Boise State University, Boise, ID, USA. ¹⁴¹Tropical Biodiversity Section, MUSE - Museo delle Scienze, Trento, Italy. ¹⁴²Department of Biology, University of Florence, Florence, Italy. ¹⁴³Department of Environmental Sciences, Central University of Jharkhand, Ranchi, India. ¹⁴⁴Faculty of Biology, Geobotany, University of Freiburg, Freiburg im Breisgau, Germany. ¹⁴⁵National Forest Centre, Forest Research Institute Zvolen, Zvolen, Slovakia. ¹⁴⁶Université de Lorraine, AgroParisTech, Inra, Silva, Nancy, France. ¹⁴⁷Center for Biodiversity Dynamics in a Changing World (BIOCHANGE), Department of Bioscience, Aarhus University, Aarhus, Denmark. ¹⁴⁸Departamento de Biología, Universidad de la Serena, La Serena, Chile. ¹⁴⁹Centro de Ciências Biológicas e da Natureza, Universidade Federal do Acre, Rio Branco, Acre, Brazil. ¹⁵⁰Guyana Forestry Commission, Georgetown, French Guiana. ¹⁵¹Faculty of Science, Universiti Brunei Darussalam, Bandar Seri Begawan, Brunei Darussalam. ¹⁵²Plant Systematic and Ecology Laboratory, Department of Biology, Higher Teachers' Training College, University of Yaoundé, Yaoundé, Cameroon. ¹⁵³Departamento de Ecologia, Universidade Federal do Rio Grande do Norte, Natal, Brazil. ¹⁵⁴Section for Ecoinformatics & Biodiversity, Department of Bioscience, Aarhus University, Aarhus, Denmark. ¹⁵⁵Faculty of Forestry and Wood Sciences, Czech University of Life Sciences, Prague, Czech Republic. ¹⁵⁶Systems Ecology, Free University Amsterdam, Amsterdam, The Netherlands. ¹⁵⁷Iwokrama International Centre for Rainforest Conservation and Development (IIC), Georgetown, French Guiana. ¹⁵⁸Botanical Garden of Ural Branch of Russian Academy of Sciences, Ural State Forest Engineering University, Ekaterinburg, Russia. ¹⁵⁹LINCGlobal, Museo Nacional de Ciencias Naturales, CSIC, Madrid, Spain. ¹⁶⁰Systematic Botany and Functional Biodiversity, Institute of Biology, Leipzig University, Leipzig, Germany. ¹⁶¹Silviculture Research Institute, Vietnamese Academy of Forest Sciences, Hanoi, Vietnam. ¹⁶²Cirad, UMR-AMAP, CNRS, INRA, IRD, Université de Montpellier, Montpellier, France. ¹⁶³Centre for the Research and Technology of Agro-Environmental and Biological Sciences, CITAB, University of Trás-os-Montes and Alto Douro, UTAD, Vila Real, Portugal. ¹⁶⁴Agricultural High School, Polytechnic Institute of Viseu, Viseu, Portugal. ¹⁶⁵Environmental Studies and Research Center, University of Campinas, UNICAMP, Campinas, Brazil. ¹⁶⁶Department of Forest and Wood Science, University of Stellenbosch, Stellenbosch, South Africa. ¹⁶⁷Key Laboratory of Tropical Biological Resources, Ministry of Education, School of Life and Pharmaceutical Sciences, Hainan University, Haikou, China. ¹⁶⁸Division of Forestry and Natural Resources, West Virginia University, Morgantown, WV, USA. ¹⁶⁹Manaaki Whenua—Landcare Research, Lincoln, New Zealand. ¹⁷⁰Department of Wetland Ecology, Institute for Geography and Geoecology, Karlsruhe Institute for Technology, Karlsruhe, Germany. ¹⁷¹Centre for Agricultural Research in Suriname (CELOS), Paramaribo, Suriname. ¹⁷²Tropenbos International, Wageningen, The Netherlands. ¹⁷³Polish State Forests, Coordination Center for Environmental Projects, Warsaw, Poland. ¹⁷⁴Programa de Pós-graduação em Biologia Vegetal, Instituto de Biologia, Universidade Estadual de Campinas, Campinas, Brazil. ¹⁷⁵Spatial Ecology and Conservation Laboratory, School of Forest Resources and Conservation, University of Florida, Gainesville, FL, USA. ¹⁷⁶Flamingo Land Ltd, Kirby Misperton, UK. ¹⁷⁷Centro Agricoltura, Alimenti, Ambiente, University of Trento, San Michele all'Adige, Italy. ¹⁷⁸Wild Chimpanzee Foundation, Liberia Office, Monrovia, Liberia. ¹⁷⁹Centro de Modelación y Monitoreo de Ecosistemas, Universidad Mayor, Santiago, Chile. ¹⁸⁰Laboratorio de Biometria, Universidad de La Frontera, Temuco, Chile. ¹⁸¹Faculty of Environmental Sciences and Natural Resource Management, Norwegian University of Life Sciences, Ås, Norway.

METHODS

We quantified the relative abundance of tree symbiotic guilds across >1.1 million forest census plots combined in the GFBi database, an extension of the plot-based GFB database³¹. The GFBi database consists of individual-based data that we compiled from all the regional and national GFBi forest-inventory datasets. The standardized GFBi data frame (that is, tree list) comprises tree identifier (ID) (a unique number assigned to each individual tree); plot ID (a unique string assigned to each plot); plot coordinates, in decimal degrees of the WGS84 datum; tree size, in diameter-at-breast-height; trees-per-hectare expansion factor; year of measurement; dataset name (a unique name assigned to each forest inventory dataset); and binomial species names of trees.

We checked all species names from different forest inventory datasets for errors in three steps. First, we extracted scientific names from original datasets, and kept only the names of genus and species (authority names are removed). Next, we compiled all the species names into five general species lists (one for each continent). Finally, we verified individual species names against 23 online taxonomic databases using the 'taxize' package of the R programming language³². We assigned each morphospecies a unique name that comprised the genus, the string 'spp', followed by the dataset name and a unique number for that species. For example, 'Picea sppCNI1' and 'Picea sppCNI2' represent two different species under the genus *Picea*, observed in the first Chinese dataset (CNI).

We derived plot-level abundance information in terms of species-abundance matrices. Each species-abundance matrix consisted of the number of individuals by species (column vectors) within individual sample plots (row vectors). In addition, key plot-level information was also added to the matrices, including plot ID, dataset name, plot coordinates, the year of measurement and basal area (that is, the total cross-sectional areas (in m²) of living trees per one hectare of ground area).

Tree genera were assigned to a plant family using a plant taxonomy lookup table generated by W. Cornwell (hosted on Github, <https://github.com/traitecoevo/taxonomylookup>), which uses the accepted taxonomy from 'The Plant List' (<http://www.theplantlist.org/>). The majority (96.5%) of genera of the species in the GFBi were successfully matched to family; for those that could not be assigned, we manually checked the genus and species in the GFBi against synonyms from The Plant List. Of the 1,038 mismatches that remained after automated assignment to families, an additional 440 genera were assigned to family either by updating older genera and species names with their more-recent synonyms or by correcting obvious misspellings. The remaining 598 entries that could not be matched to family were excluded from further analysis.

We used a taxonomically informed approach to assign symbiotic states to plant species from the GFBi. Plant species were assigned to one of five symbiotic guilds: ectomycorrhizal, arbuscular mycorrhizal, ericoid mycorrhizal, weakly arbuscular mycorrhizal or non-mycorrhizal (AMNM) or N-fixer (Supplementary Table 1). Although we did not model the relative abundance of ericoid mycorrhizal trees (owing to their rarity), we have included a map of their relative abundance from our grid (Supplementary Fig. 1). We also include the full species list as Supplementary Data; this list includes the columns used to assign species to guilds. We also include a list of families and genera assigned to all guilds except the arbuscular mycorrhizal guild (Supplementary Tables 2–5), with notes for cases of species of individual genera that were assigned to two guilds simultaneously (for example, *Alnus* is an N-fixer and ectomycorrhizal) or for cases in which species from individual genera were split between two different guilds (for example, some *Pisonia* sp. are AMNM and some are ectomycorrhizal). An arbuscular mycorrhizal summary table is excluded from the Supplementary Tables for length considerations; this information is available as Supplementary Data (file name 'SymbioticGuildAssignment.csv').

The taxonomy of species in our inventory was compared with recently published literature on the evolutionary history of mycorrhizal symbiosis^{7,33} and nitrogen fixation^{34–37}. For most species, symbiotic status could be reliably assigned at the genus (for example, *Dicymbe*) or family level (for example, Pinaceae). For the few groups for which status was unreliable or variable within a genus (for example, *Pisonia*), we conducted additional literature searches.

We assigned species to the ectomycorrhizal category in three stages: first, at the family level (for example, Pinaceae); then, at the genus level (for example, *Dicymbe*); and, finally, by using literature searches for genera for which the status was unclear (for example, in the genus *Pisonia* some species are arbuscular mycorrhizal and others are ectomycorrhizal). We used a published list³⁸ to sort species into the appropriate guild. For the genus *Acacia*, we followed previous work⁷ by assuming that only endemic Australian species associate with ectomycorrhizal fungi (we sorted *Acacia* species according to provenance using <http://worldwidewattle.com/>).

The AMNM category grouped all genera of terrestrial, non-epiphytic plants that either lack arbuscular mycorrhizal fungi or have low or inconsistent records

of arbuscular mycorrhizal fungi colonization of roots. For example, although there are some published records of arbuscular mycorrhizal fungi colonization in the roots of plants of the Proteaceae family, these records are inconsistent and colonization is generally low. Further, as Proteaceae are associated with a non-mycorrhizal root morphology (the cluster or proteoid root system) that allows them to access otherwise unavailable forms of soil nutrients³⁹, we placed the entire family within AMNM. The family Urticaceae (which we also characterized as AMNM) was problematic—early successional species from tropical forests, such as those in the genus *Cecropia*, have records of both low and absent arbuscular mycorrhizal fungi colonization⁴⁰. Our approach was to use the most broadly inclusive categorization for AMNM plants.

N-fixer status was assigned at the genus level, using previously compiled databases of global symbiotic N₂ fixation^{34–37}. Given that symbiotic N₂ fixation with rhizobial or *Frankia* bacteria has evolved only in four orders (Rosales, Cucurbitales, Fabales and Fagales)⁴¹, all species outside of this nitrogen-fixing clade were assigned non-fixing status. Some species could not be assigned an N-fixer status because they were typed to a higher taxonomic level (for example, family) that is ambiguous from the perspective of N-fixer status. We recorded when our assignment of N-fixer status was based on phylogenetic criteria, but where symbiotic nitrogen fixation is evolutionarily labile. Because these cases are more likely to be mis-assigned, we excluded them from the nitrogen-fixation category. The N-fixer group contains species that are colonized by arbuscular mycorrhizal fungi (for example, most genera from Leguminosae) and others that are colonized by ectomycorrhizal fungi (for example, *Alnus* sp.).

Most plant species form arbuscular mycorrhizal symbioses, the basal symbiotic state relative to the later-derived ectomycorrhizal and nitrogen-fixing symbioses. Furthermore, many ectomycorrhizal and nitrogen-fixing plants maintain the ability to form arbuscular mycorrhizal symbioses. Thus, a tree species is most likely to be arbuscular mycorrhizal if it does not form associations with another symbiotic guild (or forgoes root symbiosis entirely), as evidenced by its inclusion in exhaustive databases of plant symbiotic state^{7,33–37,40}. In keeping with other large-scale studies in the field³³, we assigned tree species from the GFBi database an arbuscular-mycorrhizal-exclusive state if they belonged to taxa that were not matched to ectomycorrhizal, ericoid mycorrhizal, AMNM or N-fixer symbioses. Thus, the arbuscular mycorrhizal and N-fixer groups in our dataset are non-overlapping, despite the fact that most N-fixers also associate with arbuscular mycorrhizal fungi.

The proportions of tree basal area and tree individuals were aggregated to a 1°-by-1° grid by taking the weighted average of the plot-level proportions (Supplementary Table 6). This resulted in a total of 2,768 grid cells, each with a score for the proportional abundance of ectomycorrhizal, arbuscular mycorrhizal, N-fixer, ericoid mycorrhizal and AMNM trees. We calculated two measures of relative abundance for each symbiotic guild: the proportion of tree stems and the proportion of tree basal area. Because the measurements are highly correlated with one another (Supplementary Fig. 2), we chose to model only the proportion of total tree basal area, which should scale more closely to proportion of tree biomass as it accounts for differences in size among individual stems. Additionally, we quantified variability among plots within each grid cell by calculating the weighted standard deviation across the grid (Supplementary Information, Supplementary Figs. 3, 4).

To identify the key factors that structure symbiotic distributions, we assembled 70 global predictor layers: 19 climatic indices (relating to annual, monthly and quarterly temperature and precipitation variables), 14 soil chemical indices (relating to total soil nitrogen density, microbial nitrogen, C:N ratios and soil phosphorus fractions, pH and cation exchange capacity), 5 soil physical indices (relating to soil texture and bulk density), 26 vegetative indices (relating to leaf area index, total stem density, enhanced vegetation index means and variances) and 5 topographic variables (relating to elevation and hillshade) (Supplementary Table 7). Because decomposition is the dominant process by which soil nutrients become available to plants, we generated five additional layers that estimate climatic control of decomposition. We parameterized decomposition coefficients according to the Yasso07 model^{20,42}, using the following equation: $k = \exp(0.095T - 0.00014 \times T^2) \times (1 - \exp[-1.21 \times P])$, in which P and T are precipitation and mean temperature (either quarterly or annually) of a grid cell, and the constants 0.095, 0.00014 and -1.21 are parameters that were fit using a previous global study of leaf litter mass loss²⁰. Although local decomposition rates can vary considerably based on litter quality or microbial community composition⁴³, climate is the primary control at the global scale²⁰. Decomposition coefficients describe how fast different chemical pools of leaf litter lose mass over time, relative to a parameter (α) that accounts for leaf chemistry. Decomposition coefficients (k) with values of 0.5 and 2 indicate a halving and doubling of decomposition rates, respectively, relative to α (Supplementary Information, Supplementary Fig. 5).

We implemented the random-forest algorithm using the 'randomForest' package in R. Random-forest models average over multiple regression trees, each of which uses a random subset of all the model variables to predict a response. We first determined the influence and relationship of all 75 predictor layers on forest symbiotic state, and then optimized our models using a stepwise reduction in variables from least to most important. Variable importance was measured in two ways: increase in node purity and percentage increase in MSE (with values reported in Fig. 2). The increase in node purity of variable x considers the decrease in the residual sum of squares that results from splitting regression trees using variable x . The percentage increase in MSE quantifies the increase in model error as a result of randomly shuffling the order of values in the vector x . We chose to rank variables according to the increase in node purity because we found that higher increases in node purities were associated with larger effect sizes, whereas larger percentage increases in MSE were associated with more-linear responses with smaller effect sizes. Whereas our inspection of partial feature contributions is derived from univariate random-forest models, we additionally ran multivariate random forests that predict the proportional abundance of ectomycorrhizal, arbuscular mycorrhizal and N-fixer trees for each pixel. The multivariate models were run using 50 regression trees each, with the unique set of the best 4 predictor variables for each symbiotic guild in the univariate models (Fig. 2, Supplementary Table 7). Despite strong negative correlations between the proportions of ectomycorrhizal and arbuscular mycorrhizal basal area (Supplementary Fig. 22), the results from multivariate and univariate random forests are strongly correlated with one another (Supplementary Fig. 23).

Using model selection based on eliminating variables with a low increase in node purity, we removed most soil nutrient, vegetative and topographic variables from our models (Supplementary Figs. 6, 7). Our final models include the remaining 34 predictor layers with climate, decomposition and some soil physical and chemical information (Supplementary Fig. 8). To determine the parsimony of our models, we compared the coefficient of determination in models run with a stepwise reduction in the number of variables (starting with those with the lowest increase in node purity). Based on performance of the ratio of coefficient of determination in models with 4 versus 34 variables, we determined that the 4 most-important variables accounted for >85% of the explained variability (Supplementary Fig. 9). We also compared model performance visually with plots of actual versus predicted proportions of each tree symbiotic guild among continents and geographical subregions (Supplementary Fig. 10). We used the 'forestFloor' package in R to plot the partial variable response of tree symbiotic guilds to each predictor variable (Fig. 2a–c, see Supplementary Figs. 19–21 for partial plots of the partial feature contributions of all 34 variables).

To test the sensitivity of model performance and predictions, we performed cross-validation in R using the 'rfUtilities' package⁴⁴. K -fold cross-validation tests the sensitivity of model predictions to losing random subsets from the training data. For ectomycorrhizal, arbuscular mycorrhizal and N-fixer models, we ran 99 iterations that withheld 10% of the model training data. We assessed the decrease in model performance in the 99 iterations by manually calculating the coefficient of determination, which uses the following formula: $1 - \frac{\sum(\text{actual percentage basal area} - \text{predicted percentage basal area})^2}{\sum(\text{actual percentage basal area} - \text{mean actual percentage basal area})^2}$. For all symbiotic guilds, withholding 10% of the training data resulted in a mean loss in variance explained of less than 1% (Supplementary Fig. 11). This shows that our training data have sufficient redundancy to ensure that our model conclusions are robust. Similarly, to determine whether our random-forest models would make similar predictions if data were equally distributed among continents, we rarefied our aggregated grid of symbiotic states and predictor layers to an even depth. Specifically, we sub-sampled all continents—North America (including Central America and the Caribbean), South America, Europe, Asia and Oceania—to match the number of grid pixels from Africa ($n = 50$). This is a much more aggressive reduction of training data than is typically used in K -fold cross-validations, as it involves dropping ~90% of training data rather than retaining the same amount. We performed 99 iterations of rarefaction each for the three symbiotic guilds. On average, models run with the rarefied data explained about 10% less variance over the full training data (the entire predictor/response grid) than did models run with all of the training data (Supplementary Figs. 12, 13).

To avoid projecting our random-forest models outside the ranges of their training data (for example, grid cells with higher mean annual temperatures than the maximum used to fit the models), we subset a global grid of predictor layers depending on whether (1) the grid cell fell within the top 60% of land surface with respect to tree stem density¹¹ and either (2) fell within the univariate distribution of all the predictor layers from our training data and/or (3) fell within an 8-dimensional hypervolume defined by the unique set of the 4 best predictors of the relative abundance of each guild (Supplementary Fig. 14). We then projected our models across only those grid cells that met these criteria, which constitutes 46% of the global land surface and 88% of global tree stems

(Fig. 1, Supplementary Fig. 15). Model projections were made at two resolutions: 1°-by-1° and 0.5°-by-0.5° (Fig. 4). Although model validation indicates that our projections are robust, additional studies to ground-truth these predictions and identify any discrepancies would be valuable. If such discrepancies exist, they can help to fine-tune climate–symbiosis models, or identify areas in which climate might favour invasion by symbioses that have not yet evolved in or dispersed to a particular biogeographical region.

We used the following equation to estimate the percentage of global tree stems that belong to each tree symbiotic guild: $\frac{\sum_i(\text{predicted proportion of trees of guild } g \text{ in pixel } i) \times (\text{total number of tree stems in pixel } i)}{\sum_i(\text{total number of tree stems in pixel } i)}$. The proportion of tree stems and the proportion of tree basal area in each guild are highly correlated throughout the training data (Supplementary Fig. 4). The figures cited in the main text for each guild were calculated using model projections across all pixels, even those that did not meet the criteria for model projection because they fell outside the multivariate distribution of the predictor layers or had insufficient stem density. However, our estimates for the global percentage of trees occupied by each tree symbiotic guild change by <1% when using only those pixels that met our criteria for model projection.

In the main text, we state that sharp transitions between dominant symbiotic states with climate variables could lead to declines in ectomycorrhizal trees, particularly in the southern range limit of the northern boreal forests. To determine this, we projected our random-forest models for each symbiotic guild using climate-change projections over our 19 bioclimatic variables (Supplementary Table 7), including the decomposition coefficients that use temperature and precipitation values. Specifically, we considered the 2070 scenario with a relative concentration pathway of 8.5 W per m², which predicts an increase of greenhouse gas emissions throughout the twenty-first century⁴⁵. We plot the difference in the proportion of forest basal area between the projections for 2070 and projections that use current climate data (Supplementary Table 7, Supplementary Fig. 24). We qualify this prediction with the note that vegetative changes to forests are constrained by rates of mortality, recruitment and growth.

After training and cross-validating our models with GFBi data exclusively, we additionally tested whether our models accurately predicted the previously published⁴⁶ symbiotic state of Eurasian forests. We assigned symbiotic status to all of the trees in this previous publication, and aggregated plot-level data to a 1°-by-1° grid using the same methods as with the GFBi dataset (Supplementary Fig. 25). We found that—on average—our models predicted the symbiotic state in the regional dataset within 13.6% of the value of this previously published dataset (Supplementary Fig. 26). For projected maps in Fig. 4a–c, we included the previously published⁴⁶ data with the GFBi training data to increase geographical coverage throughout Eurasia.

Reporting summary. Further information on research design is available in the Nature Research Reporting Summary linked to this paper.

Data availability

Information regarding symbiotic guild assignments, model selection (including global rasters of our model projections for ectomycorrhizal, arbuscular mycorrhizal and N-fixer proportion of tree basal area) and analyses is available as Supplementary Data. The GFBi database is available upon written request at <https://www.gfbinitiative.org/datarequest>. Any other relevant data are available from the corresponding authors upon reasonable request.

- Liang, J. et al. Positive biodiversity–productivity relationship predominant in global forests. *Science* **354**, aaf8957 (2016).
- Chamberlain, S. A. & Szöcs, E. taxize: taxonomic search and retrieval in R. *F1000Res* **2**, 191 (2013).
- Brundrett, M. C. Mycorrhizal associations and other means of nutrition of vascular plants: understanding the global diversity of host plants by resolving conflicting information and developing reliable means of diagnosis. *Plant Soil* **320**, 37–77 (2009).
- Werner, G. D., Cornwell, W. K., Sprent, J. I., Kattge, J. & Kiers, E. T. A single evolutionary innovation drives the deep evolution of symbiotic N₂-fixation in angiosperms. *Nat. Commun.* **5**, 4087 (2014).
- Werner, G. D., Cornwell, W. K., Cornelissen, J. H. & Kiers, E. T. Evolutionary signals of symbiotic persistence in the legume–rhizobia mutualism. *Proc. Natl Acad. Sci. USA* **112**, 10262–10269 (2015).
- Afkhami, M. E. et al. Symbioses with nitrogen-fixing bacteria: nodulation and phylogenetic data across legume genera. *Ecology* **99**, 502 (2018).
- Tedersoo, L. et al. Global database of plants with root-symbiotic nitrogen fixation: Nod DB. *J. Veg. Sci.* **29**, 560–568 (2018).
- Hayward, J. & Hynson, N. A. New evidence of ectomycorrhizal fungi in the Hawaiian Islands associated with the endemic host *Pisonia sandwicensis* (Nyctaginaceae). *Fungal Ecol.* **12**, 62–69 (2014).
- Lambers, H., Martinoia, E. & Renton, M. Plant adaptations to severely phosphorus-impooverished soils. *Curr. Opin. Plant Biol.* **25**, 23–31 (2015).

40. Wang, B. & Qiu, Y.-L. Phylogenetic distribution and evolution of mycorrhizas in land plants. *Mycorrhiza* **16**, 299–363 (2006).
41. Soltis, D. E. et al. Chloroplast gene sequence data suggest a single origin of the predisposition for symbiotic nitrogen fixation in angiosperms. *Proc. Natl Acad. Sci. USA* **92**, 2647–2651 (1995).
42. Palosuo, T., Liski, J., Trofymow, J. & Titus, B. Litter decomposition affected by climate and litter quality—testing the Yasso model with litterbag data from the Canadian intersite decomposition experiment. *Ecol. Modell.* **189**, 183–198 (2005).
43. Bradford, M. A. et al. Climate fails to predict wood decomposition at regional scales. *Nat. Clim. Change* **4**, 625 (2014).
44. Evans, J. S. & Murphy, M. A. rfUtilities: random forests model selection and performance evaluation. R package version 2.1-3 <https://cran.r-project.org/web/packages/rfUtilities/> (2018).
45. Core Writing Team et al. (eds) *Climate Change 2014 Synthesis Report* (IPCC, Geneva, 2014).
46. Schepaschenko, D. et al. A dataset of forest biomass structure for Eurasia. *Sci. Data* **4**, 170070 (2017).

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see [Authors & Referees](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- ☒ ☐ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- ☐ ☒ A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- ☐ ☒ The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- ☐ ☒ A description of all covariates tested
- ☐ ☒ A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- ☐ ☒ A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- ☒ ☐ For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- ☒ ☐ For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- ☒ ☐ For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- ☒ ☐ Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

Data were derived from the publicly available Global Forest Biodiversity Initiative (GFBi)

Data analysis

All analyses were performed in R Studio (version 1.0.136), using the packages "raster", "randomForest", "forestFloor", "rgdal", "rgeos", and "hypervolume", "rUtilities", and "taxize" (using the Tree genera were assigned to a plant family using a plant taxonomy lookup table generated by Will Cornwell (hosted on Github <https://github.com/traitecoevo/taxonlookup>), which uses the accepted taxonomy from "The Plant List.") Raster images were exported from the Panoply (version 4.10.3). Multipanel figures were assembled in Inkscape (version 0.92.4).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The GFBi database is available upon written request at <https://www.gfbinitiative.org/datarequest>. Additionally, the symbiotic state assigned to tree species as a supplementary file, as are global rasters of our model projections for EM, AM, and N-fixer proportion of tree basal area.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☐ Life sciences ☐ Behavioural & social sciences ☒ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	11 tree individuals from >1.1 million forest inventory plots were assigned to symbiotic state using published data-bases of plant taxonomy vs. root-symbioses. Plots were aggregated to a 1 by 1 degree grid along with a total of 75 climatic, soil, topographic, and vegetative predictor layers. The random forest algorithm was used to determine the importance and influence of individual variables. We simplified our models by removing variables with low inc node purity, a measurement of variable of importance. We then present and interpret the environmental determinants of symbiotic state.
Research sample	The GFBi database consists of individual-based data that we compiled from all the regional and national GFBi forest inventory data sets. The standardized GFBi data frame, i.e. tree list, comprises tree ID, a unique number assigned to each individual tree; plot ID, a unique string assigned to each plot; plot coordinates, in decimal degrees of WGS84 datum; tree size, in diameter-at-breast-height; trees-per-hectare expansion factor; year of measurement; data set name, a unique number assigned to each forest inventory data set; and binomial scientific tree species names.
Sampling strategy	Plot-level data was aggregated to a 1 by 1 degree (lat / long) grid.
Data collection	GFBi plots cover a significant portion of the global forest extent, including some of the most unique forest conditions: (a) the northernmost (73°N, Central Siberia, Russia), (b) southernmost (52°S, Patagonia, Argentina), (c) coldest (-17°C annual mean temperature, Oimyakon, Russia), (d) warmest (28°C annual mean temperature, Palau, USA) plots, and (e) most diverse (405 tree species on the 1-ha plot, Bahia, Brazil). Plots in war-torn regions were assigned fuzzed coordinates to protect the identity of the plots and collaborators.
Timing and spatial scale	GFBi plots cover a significant portion of the global forest extent, including some of the most unique forest conditions: (a) the northernmost (73°N, Central Siberia, Russia), (b) southernmost (52°S, Patagonia, Argentina), (c) coldest (-17°C annual mean temperature, Oimyakon, Russia), (d) warmest (28°C annual mean temperature, Palau, USA) plots, and (e) most diverse (405 tree species on the 1-ha plot, Bahia, Brazil). Plots in war-torn regions were assigned fuzzed coordinates to protect the identity of the plots and collaborators.
Data exclusions	N/A
Reproducibility	N/A
Randomization	N/A
Blinding	N/A
Did the study involve field work?	☐ Yes ☒ No

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
☒	☐ Antibodies
☒	☐ Eukaryotic cell lines
☒	☐ Palaeontology
☒	☐ Animals and other organisms
☒	☐ Human research participants
☒	☐ Clinical data

Methods

n/a	Involved in the study
☒	☐ ChIP-seq
☒	☐ Flow cytometry
☒	☐ MRI-based neuroimaging